Research Article Open Access

Determination of Fatty Acid Content in Irradiated and Non-Irradiated Syrian Olive Oil During Storage

Mahfouz Al-Bachir^{1*} and Y. Koudsi²

¹Department of Radiation Technology, Atomic Energy Commission of Syria, Damascus, P.O. Box 6091, Syria ²Faculty of Science, Damascus University, Damascus, Syria

*Correspondence to:

Dr. Maĥfouz Al-Bachir Department of Radiation Technology, Atomic Energy Commission of Syria, Damascus P.O. Box 6091, Syria E-mail: ascientific@aec.org.sy

Received: April 01, 2019 Accepted: July 02, 2019 Published: July 05, 2019

Citation: Al-Bachir M, Koudsi Y. 2019. Determination of Fatty Acid Content in Irradiated and Non-Irradiated Syrian Olive Oil During Storage. *J Food Chem Nanotechnol* 5(3): 43-48.

Copyright: © 2019 Al-Bachir and Koudsi. This is an Open Access article distributed under the terms of the Creative Commons Attribution 4.0 International License (CC-BY) (http://creativecommons.org/licenses/by/4.0/) which permits commercial use, including reproduction, adaptation, and distribution of the article provided the original author and source are credited.

Published by United Scientific Group

Abstract

This study was carried out on Syrian olive oil (SOO) produced from *Olea europaea* L. cv Kaissy, to determine the influence of different doses of gamma irradiation treatments (0, 1, 2 and 3 kGy) and various storage periods (0, 6, 12, 24 and 36 months) on fatty acid (FA) content. As a result, the composition of FAs was determined as palmitic acid (C16:0) (13.11%); palmitoleic acid (C16:1) (0.90%); stearic acid (C18:0) (3.08%); oleic acid (C18:1) (71.37%); linoleic acid (C18:2) (10.31%) and linolenic acid (C18:3) (1.22%), in order of relative abundance. Gamma irradiation significantly increased (p<0.01) the saturated fatty acid (SFA) and decreased (p<0.01) the unsaturated fatty acids (USFA). These results indicated that the USFA/SFA ratio of the virgin olive oil was markedly decreased by irradiation at 1, 2 and 3 kGy. Long storage has a significant (p<0.05) effect in SFAs and USFAs in both irradiated and non-irradiated olive oil samples.

Keywords

Gamma irradiation, Fatty acids, Gas chromatography, Olive oil, Storage time

Introduction

Olive fruit (OF) and olive oil (OO) are very important foodstuffs worldwide since they are rich in nutrients and have an anti-oxidative activity which reduces the incidence of some diseases [1]. The Food and Drug Administration (FDA) of the USA reported the benefits of OF and OO on the risk of several diseases including coronary disease by consuming about 23 g of OO daily, due to the presence of monounsaturated fatty acids (MUFAs) in OO [2].

Olive fruits contains about 20% (wt./dry wt) of lipid consisting largely of USFAs [3]. The composition of OO is primarily triacylglycerols (99%) and secondary free fatty acids (FFAs), mono- & diacyglycerols, and an array of lipids such as sterols, hydrocarbons, tocopherols, pigments and aliphatic alcohols [4]. Triglycerides are the most dominant compounds in OO, which render the main physical and chemical properties of oil. On the other hand, the dominant FA in olive oil is oleic acid and a number of other FAs are present in tiny amounts [5]. The most important part in OO is the FAs that include stearic (18:0), oleic (18:1), linoleic (18:2), linolenic acids (18:3), palmitic (16:0), palmitoleic (16:1), and Myristic (14:0), However, eicosanoic and heptadecanoic acids are found in trace amounts [1,4]. The role of minor components present in OO has been taken into consideration, since these compounds are able to improve their biological action when VOO is consumed freshly [6]. Initially, the richness of MUFAs, mainly oleic acid, was considered as the main healthful property of VOO. After the observation that other nutrients rich in MUFA, as sunflower, rapeseeds and soybean, were not comparable as healthful food with VOO [7, 8].

Al-Bachir and Koudsi.

Processing of foods and foodstuffs by specific ionizing radiations including gamma irradiation improves microbiological safety and storability is one the most extensively studied technology of the 20th century [9, 10]. Ionizing radiation has been widely used to decontaminate both foods and spices [11]. Many studies have found that irradiation produce moderates in the FA profile in food rich in fat [12-14]. Britoet et al. [15] and Yilmaz and Gecel [16] showed that trans FAs were generated in ground beef treated with gamma irradiation. However, little information is available in the literature about the influence of gamma irradiation treatment on OFs or OOs. Also, in our knowledge there is no information about the effects of gamma irradiation treatment on FA components in OFs or OOs. Therefore, the objective of this study was to examine the effects of gamma irradiation treatments on the FA composition of OOs produced in Syria from local cultivar named Kaissy after irradiation treatment and storage period. Since the content of FAs as well as the ratio between USFAs and SFAs are important parameter for determination of nutritional value of certain oil.

Materials and Methods

Production of olive oil

Olive fruits of Kaissy cultivar, with good quality were harvested during 2010 growing season, from grove located at Deer Al Hajar research station, south Syrian region near Damascus. The oils from OFs were extracted using mechanical and physical processes [17]. The OO was extracted using the method described in the previous work [18]. Olive processing consisted of the following stages: milling and slowly mixed for about 30 min at 27 °C. Then, the paste mix was centrifuged at 3000 rpm for 3 min to extract the oil. Afterwards, the OOs were decanted and immediately transferred into dark glass bottles (500 ml) and stored at room temperature (20 - 25 °C) for analyzing. FA composition of OO samples were carried out immediately after irradiation, and after 6, 12, 24 and 36 months of storage.

Irradiation treatment

The OOs were packed in dark glass bottles and then irradiated with doses of 0, 1, 2 and 3 kGy, at room temperature (20 - 25 °C) and under normal atmospheric pressure, at a dose rate of 9.913 kGy h⁻¹ using a ⁶⁰CO irradiator (ROBO, Tech snab export, Moscow, Russia). The absorbed dose was monitored using alcoholic chlorobenzene dosimeters [10].

Fatty acids (FA) determination

The analysis was carried out using the method described in the previous work [18]. FA analysis of the samples which convert to methyl ester were made in the model of 17 Shimadzu gas chromatography apparatus (Shimadzu Corp., Kyoto, Japan) equipped with a flame ionization detector and a capillary column (CBP20-S25- 050, Shimadzu, Australia). The FA percentages were calculated by means of the CLASS - VP 4.3 program (Shimadzu Scientific Instruments, Inc., Columbia, MD, USA).

Statistical analysis

Three replicates of each treatment were used, and the entire assay was carried out in triplicate. The results were expressed as mean value and standard division (SD). Data regarding each parameter were analyzed using the SUPERANOVA computer package (Abacus Concepts Inc., Berkeley, CA, USA; 1998). The differences among means at p<0.05 were compared by using Fisher test [19].

Results and Discussion

Fatty acid profile of Syrian olive oil (SOO)

The fatty acid (FA) contents of VOO were measured by gas chromatography, and the obtained data of the SOO samples under the study regarding FAs content (%) were shown in tables 1, 2 and 3. Six main FAs were determined and divided into three group; saturated SFA, MUFA and polyunsaturated fatty acids (PUFA). Two main FAs composed in SFA group: palmitic acid (C16:0) (13.11%), and stearic acid (C18:0) (3.08%). MUFA was the most dominant group of FAs in the analyzed samples. Within this group, the oleic acid (C18:1) remained the most dominant, showing a mean amount of 71.37%.

Table 1: Changes of saturated fatty acids (palmitic (C16:0) and stearic (C18:0) content (%) on Syrian olive oil (SOO) during gamma irradiation and storage period.

Treatment	Control	1 KGY	2 KGY	3 KGY	P-level
C16:0					
0	13.11 ± 0.14 ^{bC}	14.36 ± 0.05 ^{aB}	14.40 ± 0.13 ^{aAB}	14.23 ± 0.15 ^{aA}	slok
6	13.21 ± 0.01 ^{bC}	14.21 ± 0.16 ^{aB}	14.14 ± 0.30 ^{aB}	14.15 ± 0.14 ^{aA}	slok
12	13.96 ± 0.17 ^{cA}	14.92 ± 0.34 ^{bA}	14.52 ± 0.14 ^{aA}	14.46 ± 0.03 ^{aA}	slok
24	13.51 ± 0.05 ^{cB}	14.64 ± 0.17 ^{aB}	14.21 ± 0.08 ^{bB}	14.49 ± 0.36 ^{aA}	sjoje
36	13.87 ± 0.04 ^{dA}	15.11 ± 0.04 ^{aA}	14.55 ± 0.02 ^{bA}	14.32 ± 0.13 ^{cA}	slok
P-level	\$08	**	*	NS	
C18:0	•				
0	3.08 ± 0.29^{aA}	3.07 ± 0.38^{aA}	2.21 ± 0.40 ^{bC}	2.28 ± 0.30^{bB}	
6	1.96 ± 0.06 ^{bC}	2.31 ± 0.21 ^{aC}	2.13 ± 0.20 ^{abC}	2.05 ± 0.13 ^{abC}	*
12	2.66 ± 0.01^{dB}	2.98 ± 0.01 ^{aA}	2.89 ± 0.02 ^{bA}	2.77 ± 0.07^{cA}	slok
24	2.36 ± 0.22 ^{aB}	2.51 ± 0.11 ^{aB}	2.35 ± 0.08 ^{aBC}	2.27 ± 0.24 ^{aB}	NS
36	2.39 ± 0.02^{dB}	2.68 ± 0.05^{aB}	2.62 ± 0.01 ^{bAB}	2.47 ± 0.01 ^{cAB}	slok
P-level	\$ek	**	*	100	

abc Significant difference between irradiation treatments are presented with different superscript (* p<0.05, **p<0.01).</p>

NS: not significant.

The total SFAs, total USFA), and the ratio USFA/SFA of the SOO are presented in table 4. The results showed that the total SFA of SOO was found in a low amount (16.19%). While the amount of USFA and TUSFA/TSFA of the same sample was found in high level 83.81% and 5.18, respectively.

The value of oleic acid obtained in the present study from Kaissy variety of olive was comparable to the standard values

 $^{^{}ABC}$ Significant difference between storage periods are presented with different superscript (* p<0.05, ** p<0.01).

of oleic acid reported for olive oil in the literature [20]. The presence of high content of the essential oleic acid suggests that olive oil is highly nutritious. Palmitoleic acid (C16:1) (0.90%) was the second MUFA. PUFA was the third group of FAs present in SOO samples. Within this group, linoleic acid (C18:2) was the most dominant compound with average of 10.31%. Linolenic acid (C18:3) was, in order of relative abundance, the second most dominant analyst from the PUFA, with a mean concentration of 1.22%. It is well known fact that the oils of plant origin contain very small stearic acid fraction. The FA profiles of SOO revealed in our study were in synchronization with a number of already published reports on oil extracted from different commodities [21-25].

Table 2: Changes of monounsaturated fatty acids (palmitoleic (C16:1) and oleic (C18:1)) content (%) on Syrian olive oil (SOO) during gamma irradiation and storage period.

Treatment	Control	1kGy	2kGy	3kGy	P-level	
Storage time (Months)						
C16:1						
0	$0.90 \pm 0.02^{\text{bAB}}$	1.23 ± 0.08 aA	1.15 ± 0.06^{aBB}	1.15 ± 0.02 ^{aA}	slok	
6	0.95 ± 0.10 ^{bA}	1.30 ± 0.04 ^{aA}	1.26 ± 0.12 ^{aA}	1.21 ± 0.13 ^{aA}	slok	
12	0.87 ± 0.04^{cAB}	1.32 ± 0.16^{aA}	1.09 ± 0.01^{bB}	0.97 ± 0.09bcB	sjojs	
24	0.83 ± 0.08^{cB}	1.23 ± 0.04 ^{aA}	1.04 ± 0.05 ^{bB}	1.01 ± 0.05 ^{bB}	slok	
36	0.91 ± 0.01 ^{dAB}	1.25 ± 0.03 ^{aA}	1.09 ± 0.02 ^{bB}	1.00 ± 0.03cB	slok	
P-level	*	NS	*	*		
C18:1						
0	71.37 ± 0.17 ^{aB}	68.79 ± 0.33°C	70.57 ± 0.23 ^{bB}	70.94 ± 0.12 ^{bA}	slok	
6	73.62 ± 0.27 ^{aA}	69.96 ± 0.12 ^{cB}	71.10 ± 0.10 ^{bA}	71.55 ± 0.51 ^{bA}	slok	
12	71.00 ± 0.18 ^{aB}	$67.67 \pm 0.07^{\text{dD}}$	68.82 ± 0.14°C	69.33 ± 0.24 ^{bB}	slok	
24	73.26 ± 0.38 ^{aA}	70.41 ± 0.18 ^{cA}	71.21 ± 0.37 ^{bA}	71.44 ± 0.52 ^{bA}	sjojs	
36	73.08 ± 0.05 ^{aA}	$70.16 \pm 0.07^{\text{dAB}}$	70.91 ± 0.05 ^{cAB}	71.48 ± 0.11 ^{bA}	sjojs	
P-level	**	**	**	**		

abe Significant difference between irradiation treatments are presented with different superscript

Olive oil is nutritionally considered one of the best salad vegetable oil due to the highest MUFA content (75-77%), which is mainly due to the predominant presence of oleic acid [26]. The fatty acids concentration in olives vary depending on the olive variety, the ecological and environmental conditions of the location where the olives are produced, and the cultivation methods that are employed [27]. Samia Dabbou et al. [28] reported that fatty acids composition of Tunisian olive oils, palmitic acid was within the range of 9.45-11.25%, stearic acid from 2.6-2.95%, oleic acid from 66.21-72.81%, and linoleic acid from 10.92-14.92%. Aparico and Luna [29] reported that the contents of the major FAs in olive oil from Coratina, Koroneiki and Picual varieties varied between 78.1-80.3% oleic, 9.7-11.6% palmitic, 4.8-5.7% linoleic, 2.2-2.4%

stearic and 0.4-0.8% linolenic acids.

The MUSFA have great importance because of their nutritional impact and their effects on oxidative stability of oils [30]. High PUFA percentage contributes to a healthier product, however it is very important to be aware of the possible reduced storage stability and problems related to fat oxidation [31].

Effect of gamma irradiation on FA profile of Syrian olive oil (SOO)

The effect of gamma irradiation doses (0, 1, 2 and 3 kGy) in FA content of Syrian olive oil (SOO) was determined. Data presented in tables 1 - 4, showed that the single FA content, total SFAs, total USFAs, and the ratio USFA/SFA of the olive oil of Kaissy cultivar were significantly (p<0.05) changed by gamma irradiation. All used doses of gamma irradiation (1, 2 and 3 kGy) increased significantly (p<0.01) the percentage of palmitic acid (C16:0). While, only the higher doses of gamma irradiation (2 and 3 kGy) significantly (p<0.01) decreased the percentage of stearic acid (C18:0) (Table 1). Regarding the MUSFAs, 1, 2 and 3 kGy doses of gamma irradiation significantly (p<0.01) increased the percentage of palmitoleic acid (C16:1) and decreased (p<0.01) the percentage of oleic acid (C18:1) (Table 2). Finally, used doses of gamma irradiation significantly (p<0.01) increased the percentage of linoleic acid (C18:2) and decrease (p<0.05) the percentage of Linolenic acid (C18:3) (Table 3). In addition, gamma irradiation significantly (p<0.01) increase the total SFAs and decreased

Table 3: Changes of polyunsaturated fatty acids (linoleic (C18:2) and Linolenic (C18:3) acids) content (%) on Syrian olive oil (SOO) during gamma irradiation and storage period.

Treatment	Control	1kGy	2 kGy	3 kGy	P-level
Storage time	e (Months)				
C18:2					
0	10.31 ± 0.07 ^{dA}	11.52 ± 0.01 ^{aA}	10.89 ± 0.10 ^{bA}	10.64 ± 0.11 ^{cA}	slok
6	9.38 ± 0.10 ^{dC}	11.19 ± 0.30 ^{aB}	10.54 ± 0.07 ^{bB}	10.17 ± 0.13 ^{cC}	stok
12	9.60 ± 0.05^{cB}	10.95 ± 0.23 ^{aB}	10.63 ± 0.11 ^{bB}	10.42 ± 0.16 ^{bB}	stok
24	9.35 ± 0.12 ^{bC}	10.60 ± 0.25 ^{aC}	10.52 ± 0.19 ^{aB}	10.14 ± 0.38aCD	slok
36	9.19 ± 0.02^{dD}	10.23 ± 0.06 ^{aD}	10.16 ± 0.02 ^{bC}	10.02 ± 0.01 ^{cD}	**
P-level	808	88	**	*	
C18:3				,	
0	1.22 ± 0.32^{aB}	1.04 ± 0.11 ^{abB}	0.79 ± 0.02^{bB}	$0.77 \pm 0.01^{\text{bBC}}$	*
6	0.89 ± 0.01 ^{aC}	1.02 ± 0.24 ^{aB}	0.83 ± 0.20 ^{aB}	0.88 ± 0.15 ^{aB}	NS
12	1.91 ± 0.01 ^{cA}	2.15 ± 0.06 ^{aA}	2.06 ± 0.05 ^{bA}	2.00 ± 0.01 ^{bA}	**
24	0.69 ± 0.03^{aCD}	0.60 ± 0.06^{bC}	0.67 ± 0.04 ^{abB}	0.66 ± 0.03 ^{abC}	*
36	0.56 ± 0.02^{bD}	0.57 ± 0.08 ^{bC}	0.67 ± 0.01 ^{aB}	0.71 ± 0.04 ^{aC}	*
P-level	808	808	state	808	

abe Significant difference between irradiation treatments are presented with different superscript

^{(*} p<0.05, **p<0.01).

 $^{^{\}mbox{\scriptsize ABC}}$ Significant difference between storage periods are presented with different superscript

^{(*} p<0.05, ** p<0.01).

NS: not significant.

^{(*} p<0.05, **p<0.01).

 $^{^{\}mbox{\scriptsize ABC}}$ Significant difference between storage periods are presented with different superscript

^{(*} p<0.05, ** p<0.01).

NS: not significant.

(p<0.05) the total USFAs and the ratio of TUSFA to TSFA. In the present study, this ratio was 5.18 prior to irradiation of SOO decreasing to 4.74, 5.02 and 5.06 after irradiation of oil with 1, 2 and 3 kGy, respectively (Table 4).

Table 4: Changes of total saturated fatty acids (SFA), total unsaturated fatty acids (USFA), (TUSFA/SFA), and (PUSFA/SFA) on Syrian olive oil (SOO) during gamma irradiation and storage period.

Treatment	Control	1kGy	2kGy	3kGy	P-level	
Storage time (Months)						
SFA						
0	16.19 ± .0.26 ^{bB}	17.43 ± 0.36 ^{aAB}	16.61 ± 0.28 ^{bB}	16.50 ± 0.19 ^{bBC}	xiok	
6	15.16 ± 0.06 ^{bD}	16.52 ± 0.31 ^{aC}	16.27 ± 0.17 ^{aC}	16.20 ± 0.25 ^{aC}	ilak	
12	16.61 ± 0.16 ^{cA}	17.91 ± 0.35 ^{aA}	17.41 ± 0.16 ^{bA}	17.23 ± 0.10 ^{bA}	stok	
24	15.88 ± 0.17°C	17.15 ± 0.07 ^{aB}	16.56 ± 0.16 ^{bBC}	16.76 ± 0.14 ^{bB}	stok	
36	16.27 ± 0.06 ^{dB}	17.79 ± 0.05 ^{aA}	17.18 ± 0.01 ^{bA}	16.79 ± 0.13 ^{cB}	stote	
P-level	**	808	**	**		
USFA						
0	83.81 ± 0.26 ^{aC}	82.57 ± 0.36 ^{bC}	83.39 ± 0.28 ^{aB}	83.50 ± 0.19 ^{aAB}	slok	
6	84.84 ± 0.06 ^{aA}	83.48 ± 0.30 ^{hA}	83.73 ± 0.17 ^{bC}	83.80 ± 0.24 ^{bA}	slok	
12	83.39 ± 0.16 ^{aD}	82.09 ± 0.35 ^{cD}	82.59 ± 0.16 ^{bA}	82.71 ± 0.14 ^{bC}	slok	
24	84.12 ± 0.17 ^{aB}	82.85 ± 0.07 ^{cB}	83.45 ± 0.16 ^{bBC}	83.25 ± 0.14 ^{bB}	slok	
36	83.73 ± 0.06 ^{aC}	82.21 ± 0.05 ^{dD}	82.82 ± 0.01 ^{cA}	83.21 ± 0.12 ^{bB}	slok	
P-level	**	sjojs	\$6\$	**		
USFA/SF	Ά					
0	5.18 ± 0.10^{aC}	4.74 ± 0.12 ^{bBC}	5.02 ± 0.10^{aB}	5.06 ± 0.07^{aAB}	slok	
6	5.60 ± 0.03 aA	5.05 ± 0.11^{bA}	5.15 ± 0.07^{bA}	5.17 ± 0.09 ^{bA}	slok	
12	5.02 ± 0.06^{aD}	4.59 ± 0.11 ^{cC}	4.74 ± 0.05 ^{bC}	4.80 ± 0.03 ^{bC}	sjojs	
24	5.30 ± 0.17^{aB}	4.83 ± 0.02 ^{cB}	5.04 ± 0.06^{bAB}	4.97 ± 0.05 ^{bB}	slok	
36	5.15 ± 0.02^{aC}	4.62 ± 0.02°C	4.82 ± 0.002 ^{bC}	4.96 ± 0.04^{bB}	slok	
P-level	**	slok	**	**		
PUSFA/S	FA					
0	0.71 ± 0.03 ^{aA}	0.72 ± 0.02^{aA}	0.70 ± 0.02^{aB}	0.69 ± 0.02^{aB}	NS	
6	0.68 ± 0.01^{bB}	0.74 ± 0.03 ^{aA}	0.70 ± 0.02^{bB}	$0.68 \pm 0.01^{\mathrm{bB}}$	stots	
12	$0.69 \pm 0.01^{\text{bAB}}$	0.73 ± 0.03^{aA}	0.73 ± 0.01^{aA}	0.72 ± 0.01^{abA}	*	
24	0.63 ± 0.01^{bC}	0.65 ± 0.01^{bB}	0.68 ± 0.004 ^{aC}	0.64 ± 0.02^{bC}	alank	
36	0.60 ± 0.004^{bD}	0.61 ± 0.004 ^{bC}	$0.63 \pm 0.002^{\mathrm{aD}}$	0.64 ± 0.01^{aC}	ilak	
P-level	**	101	**	**		

abc Significant difference between irradiation treatments are presented with different superscript (* p<0.05, **p<0.01).

NS: not significant.

The value of polyunsaturated fatty acids to saturated fatty acids (P/S) indexes of tested SOO are shown in table 4. The P/S index of the investigated SOO varied depending on the treatments. The P/S indexes of the analyzed SOO samples (irradiated at 0, 1, 2 and 3 kGy and stored for 0, 6, 12, 24 and 36 months) ranged from 0.60 and 0.74. The P/S index value of the control sample of SOO is 0.71, and this value decreased with increasing the storage time. There are no significant

differences in P/S index value between the irradiated and unirradiated olive oil samples.

The concentration of total USFAs decreased significantly (p<0.01), and the total SFAs increased (p<0.05) throughout the storage period in the SOO samples analyzed. This leads to a corresponding decrease in total USFA/total SFA (oxidation index) throughout storage period for SOO (Table 4).

Tables 1, 2 and 3 show the trend of FA contents of irradiated and un-irradiated olive oil during storage. There were significant (p<0.05) differences in the single fatty acid composition, total SFAs, total USFAs, and the ratio USFA/SFA of irradiated and un-irradiated SOO samples stored for 0, 6, 12, 24 and 36 months.

The results of this study indicate that irradiation induced decomposition of the USFAs. Free radicals generated by irradiation react with the double bonds of FAs [32, 33]. Moreover, radical such as thiyl radicals, which are generated by ionizing radiation treatment during the repair of any radical, may interact with USFAs [34]. Furthermore, the cis configuration is less stable the trans configuration [35, 36]. Similar results were previously observed [37, 38]. Irradiating at 5 kGy decreased total amount of USFAs and increased the total amount of SFAs in beef lipids [39]. Another study reported that the decrease in USFAs during the irradiation process of oil is mainly due to a molecular structure moderate in fatty acids [40]. The ratio of USFA/SFA was used to predict the shelf life of hazel nuts; indication that the lower the ratio, the longer was product shelf-life [41]. On other hand, irradiation of sesame peanut and sunflower seeds at doses of 3, 6 and 9 kGy did not significantly affect the FAs percentages. However, the USFAs, SFAs and the ratio of SFAs to USFAs (TU/TS) were changed upon irradiation [42].

The relationship between PUSFAs and SFAs content is expressed as P/S index. This value is very important parameter for evaluation the nutritional value of certain oil. Oils and fats with higher value of P/S index than one is considered to have high nutritional value [26].

The behavior of the FAs during storage in SOO reported in our study was found to be in harmony with Sandchez-Bel et al. [43] and Jubeen et al. [25]. However, the individual FAs percentage of all analyzed SOO samples falls within the recommended International Oil Council [20].

Conclusion

The irradiation doses 0, 1, 2 and 3 kGy and storage period 0, 12, 24 and 36 months applied to Syrian olive oil (SOO) induce significant statistical differences (p<0.05) in FA content. It was found that, for all analyzed samples, the palmitic acid (C16:0) ranged from 13.11 to 15.11%; stearic acid (C18:0) ranged from 1.96 to 3.08%; palmitoleic acid (C16:1) ranged from 0.83% to 1.32; oleic acid (C18:1) ranged from 67.67 to 73.62%; linoleic acid (C18:2) 9.19 to 11.52%; and Linolenic acid (C18:3) varied from 0.56 to 2.15%. These fall within the recommended International Oil Council [20] for olive oils which indicate that: palmitic acid (7.50-20.00); stearic acid

 $^{^{\}rm ABC}$ Significant difference between storage periods are presented with different superscript (* p<0.05, ** p<0.01).

(0.50-5.00%); palmitoleic acid (0.30 - 3.50); oleic acid (55.00 - 83.00%); and linoleic acid (2.50 - 21.00%). Therefore, this study supports the use of gamma irradiation (up to 3 kGy) as safety treatment for SOO (stored up to 36 months) and calls for further investigations to elucidate its influence on the other chemical and physical characteristics and constituents of the oil.

Acknowledgements

The authors are grateful wish to the Director General of the Atomic Energy Commission of Syria (AECS) and the staff of food irradiation group.

Declaration of Interest

The authors report no conflicts of interest. The author alone is responsible for the content and writing of the manuscript.

Reference

- Ghanbari R, Anwar F, Alkharfy KM, Gilani AH, Saari N. 2012. Valuable nutrients and functional bioactives in different parts of olive (Olea euroaea L.) - a review. Int J Mol Sci 13(3): 3291-3340. https://doi. org/10.3390/ijms13033291
- Food and Drug Administration (FDA). 2004. Monounsaturated fatty acids from olive oil and coronary heart disease. In health claim petition docket No. 2003Q-0559, Rome, Italy.
- Unal K, Nergiz C. 2003. The effect of table olive preparing methods and storage on the composition and nutritive value of olives. *Grasas y Aceites* 54(1): 71-76. https://doi.org/10.3989/gya.2003.v54.i1.280
- Sarolic M, Gugic M, Marijanovic Z, Suste M. 2014. Virgin olive oil and nutrition. Food Health and Disease, Scientific-Professional Journal of Nutrition and Dietetics 3(1): 38-43.
- Viola P, Viola M. 2009. Virgin olive oil as fundamental nutritional component and skin protector. *Clin Dermatol* 27(2): 159-165. https://doi.org/10.1016/j.clindermatol.2008.01.008
- Bulotta S, Celano M, Lepore SM, Montalcini T, Pujia A, et al. 2014. Beneficial effect of olive oil phenolic components oleuropein and hydroxyrosol: focus on protection against cardiovascular and metabolic disease. J Transl Med 12: 219. https://doi.org/10.1186/s12967-014-0219-9
- Harpe CR, Edwards MC, Jacobson TA. 2006. Flasseed oil supplementation does not affect plasma lipoprotein concentration or particle size in human subjects. J Nutr 136(11): 2844-2848. https://doi. org/10.1093/jn/136.11.2844
- Aguilera CM, Mesa MD, Ramirez-Tortosa MC, Nestares MT, Ros E, et al. 2004. Sunflower oil does not protect against LDL oxidation as virgin olive oil does in patients with peripheral vascular disease. *Clin Nutr* 23(4): 673-681. https://doi.org/10.1016/j.clnu.2003.11.005
- Al-Bachir M. 2013. Microbiological load and quality characteristics of irradiated chicken meat. Arab Golf Journal of Scientific Research 31(1): 59-67.
- Al-Bachir M. 2004. Effect of gamma irradiation on fungal load, chemical and sensory characteristics of walnuts (*Juglans regia* L.). J Stored Prod Res 40(4): 355-362. https://doi.org/10.1016/S0022-474X(03)00030-4
- 11. Al-Bachir M, Al-Adawi MA. 2015. Comparative effect of irradiation and heating on the microbiological properties of licorice (*Glycyrrhiza glabra* L.) root powders. *Int J Radiat Biol* 91(1): 112-116. https://doi.or
- Al-Bachir M. 2014. Physicochemical properties of oil extracts from gamma irradiated almond (*Prunus amygdalus L.*). *Innov Rom Food Biotechnol* 14: 37-45.

- Al-Bachir M. 2015. Studies on the physicochemical characteristics of oil extracted from gamma irradiated pistachio (*Pistacia vera* L.). Food Chem 167: 175-179. https://doi.org/10.1016/j.foodchem.2014.06.020
- Al-Bachir M. 2015. Quality characteristics of oil extracted from gamma irradiated peanut (*Archishypogea L.*). Radiation Physics and Chemistry 106: 56-60. https://doi.org/10.1016/j.radphyschem.2014.06.026
- Britoet MS, Villavicencio ALCH, Mancini-filho J. 2002. Effects of irradiation on trans fatty acids formation in ground beef. *Radiation Physics and Chemistry* 63(3-6): 337-340. https://doi.org/10.1016/S0969-806X(01)00647-8
- Yilmaz I, Gecel U. 2007. Effects of gamma irradiation on trans fatty acid composition in ground beef. *Food Control* 18(6): 635-638. https:// doi.org/10.1016/j.foodcont.2006.02.009
- Blatchly RA, Delen Z, O'Hara PB. 2014. Making sense of olive oil: Simple experiments to connect sensory observations with the underlying chemistry. *J Chem Educ* 91(10): 1623-1630. https://doi. org/10.1021/ed300557r
- 18. Al-Bachir M, Koudsi A. 2016. Fatty acid composition of olive obtained from irradiated and non-irradiated whole fruit and fruit flesh of olives (Olea europaea L.). The Annals of the University Dunarea de Jas of Galati Fascicle VI – Food Technology 40(1): 78-89.
- Snedecor G, Cochran W. 1988. Statistical methods. The Iowa State University Press, Ames, Iowa, USA, pp 221-221.
- International Olive Council (IOOC). 2015. Trade standard applying oils and olive-pomace oil. COI/T.15/NC No 3/Rev.9 Jun 2015, pp 16.
- Amara SJ, Casal S, Pereira JA, Seabra RM, Oliveira BPP. 2003. Determination of sterol and fatty acid compositions, oxidative stability, and nutritional value of six Walnut (*Juglansregia* L.) cultivars grown in Portugal. *J Agric Food Chem* 51(26): 7698-7702. https://doi. org/10.1021/jf030451d
- Cherif A, Sebei K, Boukhchina S, Kallel H, Belkacemi K, et al. 2004.
 Kernel fatty acid and triacylglycerol composition for three almond cultivars during maturation. J Am Oil Chem Soc 81(10): 901-905. https://doi.org/10.1007/s11746-004-0999-z
- Venkatachalam M, Sathe SK. 2006. Chemical composition of selected edible nut seeds. J Agric Food Chem 54(13): 4705-4714. https://doi. org/10.1021/jf0606959
- Kornsteiner M, Wagner KH, Elmadfa I. 2006. Tocopherols and total phenolics in 10 different nut types. *Food Chem* 98(2): 381-387. https://doi.org/10.1016/j.foodchem.2005.07.033
- Jubeen F, Bhatti IB, Maqbool U, Mehbool S. 2012. Fungal incidence, aflatoxin b1, tocopherols and fatty acids dynamics in ground and tree nuts during storage at two moisture levels. *Int J Agric Biol* 14(4): 521-527.
- 26. Kostik V, Memeti S, Bauer B. 2013. Fatty acid composition of edible oils and fats. *J Hyg Eng Des* 4: 112-116.
- 27. Sakar E, Erol BA, Unver H, Celik M, Turkoglu H, et al. 2014. Determination of total olive oil and Cis-Trans fatty acids composition of Sirnak province olive genotypes at south eastern Anatolia. *Journal of Agriculture and Environmental Sciences* 3(4): 119-129. https://doi.org/10.15640/jaes.v3n4a9
- Dabbou S, Brahmi F, Dabbou S, Issaoui M, Sifi S, et al. 2011.
 Antioxidant capacity of Tunisian virgin olive oils from different olive cultivars. African Journal of Food Science and Technology 2(4): 92-97.
- Aparicio R, Luna G. 2002. Characterization of monovarietal virgin olive oil. Eur J Lipid Sci Technol 104(9-10): 614-627. https:// doi.org/10.1002/1438-9312(200210)104:9/10<614::AID-EJLT614>3.0.CO;2-L
- Aguilera MP, Beltran G, Ortega D, Fernandez A, Jimenez A, et al. 2005. Characterisation of virgin olive oil of Italian olive cultivars: 'Frantoio' and 'Leccino', grown in Adalusia. Food Chem 89(3): 387-391. https://doi.org/10.1016/j.foodchem.2004.02.046

- 31. Bryhni EA, Kjos NP, Ofstad R, Hun M. 2002. Polyunsaturated fat and fish oil diets for growing- finishing pigs: effects on fatty acid composition and meat, fat, and sausage quality. *Meat Sci* 62(1): 1-8. https://doi.org/10.1016/S0309-1740(01)00211-X
- 32. Ferreri C, Kratzsch S, Brede O, Marciniak B, Chatgilialogu C. 2005. Trans lipid formation induced by thiols in human monocytic and leukemia cells. *Free Radic Biol Med* 38(9): 1180-1187. https://doi.org/10.1016/j.freeradbiomed.2004.12.026
- 33. Chatgilialoglu C, Ferreri C. 2005. Trans lipids: the free radical path. *Acc Chem Res* 38(6): 441-448. https://doi.org/10.1021/ar0400847
- 34. Geibler C, Brede O, Reinhardt J. 2003. Cis-trans- isomerization of unsaturated fatty acids during gamma irradiation of barley grains. *Radiation Physics and Chemistry* 67(2): 105-113. https://doi.org/10.1016/S0969-806X(03)00003-3
- 35. Alfaia CM, Ribeiro PJLC, Trigo MJP, Alfaia AJI, Castro MLF, et al. 2007. Irradiation effect on fatty acid composition and conjugated linoleic acid isomers in frozen lamb meat. *Meat Sci* 77(4): 689-695. https://doi.org/10.1016/j.meatsci.2007.05.025
- Liu WH, Inbaraj S, Chen BH. 2007. Analysis and formation of trans fatty acids in hydrogenated soybean oil during heating. Food Chem 104(4): 1740-1749. https://doi.org/10.1016/j.foodchem.2006.10.069
- 37. Badr HM 2005. Chemical properties of chicken muscles and skin as affected by gamma irradiation and refrigerated storage. *Journal of Food Technology* 3(1): 1-9.

- 38. Mahmoud KA, Badr HM. 2011. Quality characteristics of gamma irradiated beef burger formulated with partial replacement of beef fat with olive oil and wheat bran fibers. *Food and Nutrition Sciences* 2(6): 655-666. https://doi.org/10.4236/fns.2011.26091
- 39. Fan X, Kays SE. 2009. Formation of trans fatty acids in beef and frankfurters due to irradiation. *J Food Sci* 74(2): C79- C84. https://doi.org/10.1111/j.1750-3841.2008.01024.x
- Arici M, Ferya AC, Umit G. 2007. Effect of gamma radiation on microbiological and oil properties of black cumin (*Nigalla sativa L.*). Grasas y Aceites 58(4): 339-343. https://doi.org/10.3989/gya.2007.v58. i4.444
- Fokou E, Achul MB, Kanscil G, Ponkal R, Fotso M, et al 2009. Chemical properties of some cucurbitaceae oil from Cameroon. Pak J Nutr 8(9): 1325-1334. https://doi.org/10.3923/pjn.2009.1325.1334
- 42. Al-Bachir M. 2017. Fatty acid contents of gamma irradiated sesame (Sesamum indicum L.) Peanut (Arachis hypogaea L.) Sunflower (Helianthus annuus L.) seeds. J Food Chem Nanotechnol 3(1): 31-37. https://doi.org/10.17756/jfcn.2017-034
- Sánchez-Bel P, Martìnez-Madrid C, Egea I, Romojaro F. 2005. Oil quality and sensory evaluation of almond (*Prunus amygdalus*) stored after electron beam processing. *J Agric Food Chem* 53(7): 2567-2573. https://doi.org/10.1021/jf040184r